

## Perspective

## Redox Metabolism in Fishes Under Thermal Stress Warrants More Attention

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**Abstract:**

Any process that maintains the redox environment in aerobes is defined as redox homeostasis. Redox homeostasis is disturbed in favor of a prognostic situation either due to increase in generation of reactive oxygen species (ROS) or due to collapse of cellular antioxidant defenses. Both conditions are greatly affected by environmental factors, and thereby, manipulate physiology of aerobes. Rise in aquatic habitat temperature induces oxidative stress in fish as a consequence of accelerated generation of ROS as by-products of elevated metabolic activities. Reactive oxygen species, if not neutralized, oxidize biomolecules present in their vicinity and impairs their biological properties and, thereby, cellular physiology. The phenomenon is referred as “oxidative stress”. Augmentation of oxidative stress is linked to various pathophysiological states reported for aerobes. Also elevation in temperature of marine or brackish water bodies results in augmentation of salinity which in turn induces oxidative stress in fish and other aquatic fauna. This may have dire consequences on fish by accelerating aging phenomenon as oxidative stress and aging are interrelated processes. Thus, it can be predicted that exposure to short intense or long gradual rise in aquatic habitat temperature due to global warming may trigger oxidative stress in aquatic fauna including fish which may affect their growth pattern, reproductive capacity, survivability and finally longevity. Fish are one of the major components of global ecosystem. Any imbalance in fish population will not only affect aquatic ecosystem but also may have dire long term consequences on global food chain. Therefore, more research must be focused not only to understand the physiological aspects related to oxidative stress in context to thermal stress for better health status in fish but also to develop new strategies for fish endurance it as thermal stress is imminent outcome of global warming.

**Keywords:** Habitat warming, Fish physiology, Fish growth, Oxidative metabolism, Thermal stress

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## Introduction

Fish belong to a paraphyletic group of organisms that consist of all gill-bearing aquatic craniates that lack limbs with digits. Fish as the major group of poikilotherms can be considered as the principal targets under rise in acute or gradual temperature of their natural habitat because these organisms lack proper physiological mechanism(s) to regulate their internal body heat in relation to changing environmental temperature (Carey and Lawson, 1973; Goldman, 1997). Consequently, their energy metabolism may be affected by increasing environmental temperature as observed in other poikilotherms (Galli and Recharads, 2012). It may result in increase in production of reactive oxygen species (ROS) either due to elevated metabolism or due to collapse of antioxidant defences, or both, which in turn may induce oxidative stress (OS) in fish by impairing functions of biomolecules as a consequence of their oxidation. OS is reported to be associated with various physiological aspects of aerobes such as growth, development and aging. Fish exhibit a great diversity in ecology and serve as a vital component in aquatic ecosystem. Also it is an important trophic level in global food chain. Therefore, increase in habitat temperature due to global warming may induce OS in fish and may hamper their growth, reproduction and aging which may indirectly affect the global ecology. This is the central focus of this perspective article.

## Oxygen Metabolism and Oxidative Stress

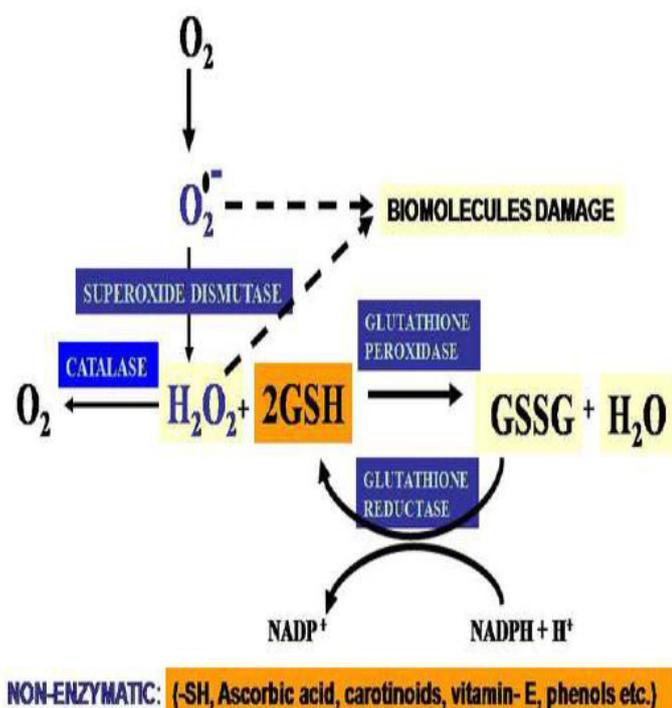
The molecular story of oxidative stress commences with consumption of  $O_2$  and oxidation of nutrient molecules to produce energy in aerobes. In the process, mitochondria play a central role. Electron transport chain of mitochondria carries electrons from universal electron donors such as  $FADH_2$  and  $NADH$  to  $O_2$  via enzyme complexes I, II, III and IV and produces ATP in complex V (Lehninger *et al.*, 2008). Incomplete reduction of oxygen molecules and subsequent leaking of electrons in complexes I and III during electron transport, results in formation of superoxide radicals (one of the ROS). Consequently, it leads to formation of other ROS such as hydrogen peroxide and hydroxyl radicals by classical Haber-Weiss reaction. Cells are equipped with efficient antioxidant defences to neutralize the ROS before they can damage biomolecules by oxidation (Halliwell and Gutteridge, 2008). However, due to decrease in efficiency of antioxidant defenses in response to internal or external insults, generation of ROS are elevated which in turn oxidize all biomolecules present in their vicinity and causes OS in cell. Under such conditions, a deviation in normal physiological functions can not be ruled out which may end up with increase in ratio between energy demand versus production, cell senescence, aging, metabolic depression, retarded growth and reproduction, and finally death may also occurs (Halliwell and Gutteridge, 2008). Therefore, careful and planned study on oxidative stress indices and redox regulatory parameters *in silico* and *in vivo* conditions will have immense importance to solve various riddles observed in several core evolutionary concepts of animal biology such as in life history tradeoffs, senescence, growth, reproduction and sexual selection in free ranging organisms (Costantini *et al.*, 2010; Paital *et al.*,

2011, 2013, 2015; Paital and Chainy, 2012; Paital and Samanta, 2013, Paital and Chainy, 2014; Paital and Chainy, 2010). Noteworthy, ROS are also reported to be useful because at lower concentrations, they mediate several signal transduction processes in cells (Passos and Zglinicki, 2006; Takano *et al.*, 2003). However, under elevated thermal stress, maintenance of the nominal amount of ROS to regulate signal transduction processes is not ensured (Passos and Zglinicki, 2006).

Aerobes are equipped with both enzymatic as well as non-enzymatic redox regulatory molecules to counteract the over production of ROS to maintain redox homeostasis. Superoxide dismutase (SOD), the first enzyme of enzymatic antioxidant defense, dismutates the toxic superoxide radicals to  $H_2O_2$  and molecular oxygen.  $H_2O_2$  is further neutralized by two cellular enzymes, catalase (CAT) and glutathione peroxidase (GPx). CAT breaks down  $H_2O_2$  to  $H_2O$  and  $O_2$  while GPx reduces  $H_2O_2$  and organic hydroperoxides to  $H_2O$  and other non-reactive metabolites at the cost of oxidation of a reduced glutathione (GSH) molecule. The oxidized glutathione is reduced back to GSH by the enzyme glutathione reductase (GR) with the help of the reduced nicotinamide adenine dinucleotide phosphate (NADPH) (Figure 1). Peroxiredoxins are a group of ubiquitous antioxidant enzymes that regulate the levels of cytokine-induced peroxides (Rhee *et al.*, 2005). The oxidized form of peroxiredoxins is non-catalytic in nature. To self recharge after reducing  $H_2O_2$ , these enzymes require thioredoxin (Rhee *et al.*, 2001). They require electrons from the reduced thioredoxin to restore their enzymatic catalytic function (Pillay *et al.*, 2009). Thioredoxins are a class of small ubiquitous redox proteins which also play a key role in removing ROS (Wollman *et al.*, 1988; Meng *et al.*, 2010). Glutaredoxins are a group of small redox enzymes which confer their antioxidant activity by reducing dehydroascorbate, peroxiredoxins and methionine sulfoxide reductase (Nordberg and Arnér, 2001). Non enzymatic antioxidant defence system comprises of small molecules such as polyphenols, carotenoids, flavonoids, ascorbic acid, vitamin E and GSH which directly scavenge ROS. Under thermal stress, the levels of redox regulatory molecules are found to be alleviated or insufficiently enhanced which fails to combat OS in organisms (Halliwell and Gutteridge, 2008).

## The Central Remark

Although not much experimental information is available on positive correlation between rise in temperature and reproduction or growth in fishes in their natural habitat, it is suggested that rise in temperature in combination with elevated  $CO_2$  level decrease the duration of nymphal stadia, the longevity and reproductive success of the aphid *Sipha flava* (Aquad *et al.*, 2012). Similarly, several authors have pointed out that increase in habitat temperature above 20-24°C, which is the optimum temperature required for the growth of aphids, can negatively influence fertility, reproduction, development, life expectancy, survival and abundance of aphids (Oliveira *et al.*, 2009; Aquad *et al.*, 2009). A common reason for the above phenomenon is provided that increase in temperature can affect the life cycle of poikilotherms directly by influencing their physiology (Flynn *et al.*, 2006). Occurrence of the above multiple processes as the consequences of climatic changes are not only



**Figure 1:** General pathway of oxidative stress metabolism.  $O_2^-$ -superoxide radicals, GSH-reduced glutathione, GSSG-oxidised glutathione.

restricted to terrestrial ecosystems but also can affect the aquatic ecosystems. For example, in aquatic environment especially in coastal marine bodies, climatic changes also modulate to elevate OS and its related metabolism as well as negatively modulate the other physiological processes such as reproduction, excretion, respiration of the inhabiting ectotherms (Lawrence and Soame, 2004; Paital and Chainy, 2010, 2012; Abele *et al.*, 2002, 2007, 2011). Increase in salinity can induce OS despite altered redox status found to be initiated to protect poikilotherms (Paital and Chainy, 2010, 2012). With altered redox regulation, fish are also found to experience OS induced by rise in habitat temperature in their natural environment. For example, in cichlid fish acar ( *Geophagus brasiliensis* ), three-spined stickleback fish (*Gasterosteus aculeatus*), Senegal sole fish (*Solea senegalensis*), environmental rise in temperature induces OS (Table 1). It has been noticed that antioxidant defence parameters changes in fish brain and muscle tissues (*Heteropneustus fossilis*) in response to air exposure due to alteration in functional capacity of electron transport chain (Paital, 2013, 2014). Temperature is known to affect oxygen content of water bodies. Therefore, molecular and cellular functions of fish due to elevated temperature as a consequence of global warming cannot be ignored.

## Conclusion

The world is projected to experience an elevation in mean global temperatures in coming years. It is presumed that an

**Table 1** Oxidative stress and redox status in natural population of some fish under high habitat temperature.

Name of the Animal	Modulator	Tissue/organ/cell fractions	OS	AOE	Reference
<i>Geophagus brasiliensis</i> (cichlid fish acar)	Comparatively high temperature in spring	Liver	TBARS, GSSG	SOD, CAT	Filho <i>et al.</i> , 2001
<i>Gasterosteus aculeatus L.</i> (three-spined stickleback Fish)	High temperature in summer with reproductive activity	Liver	TBARS	GPx	Sanchez <i>et al.</i> , 2008
<i>Lepomis macrochirus</i> (bluegill fish)	High temperature in summer	Whole animal	NA	NA	Wohlschlag and Juliano, 2003
<i>Solea senegalensis</i> (Senegal sole fish)	High temperature with heavy metal load	Liver	TBARS	CAT, GPx	Oliva <i>et al.</i> , 2012
<i>Barbus barbus L.</i> (barbell fish)	High temperature in summer	Liver	NA	SOD, CAT	Radovanovic <i>et al.</i> , 2010
<i>Barbus barbus L.</i> (barbell fish)	High temperature in summer	Muscle	NA	SOD, CAT	Radovanovic <i>et al.</i> , 2010

ROS-reactive oxygen species, OS-oxidative stress (TBARS-thiobarbituric acid reactive substances, PC-protein carbonylation), AOE-antioxidant enzymes (SOD-superoxide dismutase, CAT-catalase, GPx-glutathione peroxidase), GSSG- oxidised glutathione, NA- not analysed.  or  symbols are used to indicate decrease or increase of the parameters with the corresponding season, respectively. Parameter with no symbol ( or ) indicates “no change” in the same with respect to the season.

approximate rise in temperature of 1.4 to 5.8 °C may occur in next decades (Houghton, 2001). The rise in environmental temperature (thermal stress) of water bodies due to global warming may lead to influence redox homeostasis of aquatic organisms which in turn may be resulted into decrease in normal physiology especially growth and reproduction of poikilotherms in general and fishes in particular by augmenting OS. Therefore, understanding the physiological impacts of global warming in relation to growth and reproduction of fish in their natural environment and development of strategies to manage them are of paramount importance and should be considered as one of the major challenging areas of future research in aquaculture.

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## References

- Abele, D., Heise, K., Pörtner, H.O., and Puntarulo, S. (2002). Temperature-dependence of mitochondrial functions and production of reactive oxygen species in the intertidal mud clam *Mya arenaria* were recorded. *Exp. Biol.* **205**, 1831-41.
- Abele, D., Philipp, E., Gonzalez, P.M., and Puntarulo, S. (2007). Marine invertebrate mitochondria and oxidative stress. *Front. Biosci.* **12**, 933-946.
- Abele, D., Vazquez-Medina, J.P., and Zenteno-Savin, T. (2011). *Oxidative Stress in Aquatic Ecosystems*, first ed. Blackwell and Wiley, USA, pp 548.
- Auad, A.M., Alves, S.O., Carvalho, C.A., Silva, D.M., Resende, T.T., and Veríssimo, B.A. (2009). The impact of temperature on biological aspects and life table of *Rhopalosiphum padi* (Hemiptera: Aphididae) fed with signal grass. *Florida Entomologist.* **92**, 569-577.
- Auad, A.M., Fonseca, M.G., Resende, T.T., and Maddalena, Í.S.C.P. (2012). Effect of Climate Change on Longevity and Reproduction of *Sipha flava* (Hemiptera: Aphididae). *Florida Entomologist.* **95**, 433-444.
- Carey, F.G. and Lawson, K.D. (1973). Temperature regulation in free-swimming bluefin tuna. *Comp. Biochem. Physiol. A: Physiology*, **44**(2), 375-392.
- Costantini, D., Rowe, M., Butler, M.W., and McGraw, K.J. (2010). From molecules to living systems: historical and contemporary issues in oxidative stress and antioxidant ecology. *Funct. Ecol.* **24**, 950-959.
- Filho, D.W., Torres, M.A., Tribess, T.B., Pedrosa, R.C., and Soares, C.H.L. (2001). Influence of season and pollution on the antioxidant defenses of the cichlid fish acará (*Geophagus brasiliensis*). *Braz. J. Med. Biol. Res.* **34**, 719-726.
- Flynn, D.F.B., Sudderth, E.A. and Bazzaz, F.A. (2006). Effects of aphid herbivory on biomass and leaf-level physiology of *Solanum dulcamara* under elevated temperature and CO<sub>2</sub>. *Environ. Expt. Bot.* **56**, 10-18.
- Galli, G.L.J., and Richards, J.G. (2012). The effect of temperature on mitochondrial respiration in permeabilized cardiac fibres from the freshwater turtle, *Trachemys scripta*. *J. Therm. Biol.* **37**, 195-200.
- Goldman, K.J. (1997). Regulation of body temperature in the white shark, *Carcharodon carcharias*. *J. Comp. Physiol. B.* **167**(6), 423-429.
- Halliwell, B., and Gutteridge, J.M.C. (2008). *Free Radicals in Biology and Medicine*. Oxford University Press, New York, USA.
- Houghton, J.T., dinGy, GriGGs, dJ, noGuer, M., vAn der linden, P.J., xiAosu d, MAskell, k, and Johnson, C.A. (2001). *Climate Change 2001: The Scientific Basis*. Cambridge Univ. Press, Cambridge, UK.
- Lawrence, A.J., and Soame, J.M. (2004). The effects of climate change on the reproduction of coastal invertebrates. *Int. J. Avian. Sci.* **146**, 29-39.
- Lehninger, A.L., Nelson, D.L., and Cox, M.M. (2008). *Lehninger Principles of Biochemistry* (5th edition), W.H. Freeman & Co., New York, USA, pp. 707-764.
- Lenaz, G., Fato, R., Genova, M., Bergamini, C., Bianchi, C., and Biondi, A. (2006). Mitochondrial Complex I: structural and functional aspects. *Biochim Biophys Acta.* **1757**, 1406-1420.
- Meng, L., Wong, J.H., Feldman, L.J., PGemaux, L., and Buchanan, B.B. (2010). A membrane-associated thioredoxin required for plant growth moves from cell to cell, suggestive of a role in intercellular communication. *Proc. Nat. Acad. Sci. US.* **107**, 3900-3905.
- Nordberg, J., and Arnér, E.S. (2001). Reactive oxygen species, antioxidants, and the mammalian thioredoxin system. *Free Radic. Biol. Med.* **31**, 1287-312.
- Oliva, M., Vicente, J.J., Gravato, C., Guilhermino, L., and Galindo-Rian~o, M.D. (2012). Oxidative stress biomarkers in Senegal sole, *Solea senegalensis*, to assess the impact of heavy metal pollution in a Huelva estuary (SW Spain): Seasonal and spatial variation. *Ecotoxicol. Environ. Saf.* **75**, 151-162.
- Oliveira, S.A., Auad, A.M., Souza, B., Souza, L.S., Amaral, R.L., and Silva, D.M. (2009). Tabela de esperança de vida e de fertilidade de *Sipha flava* (Forbes) (Hemiptera, Aphididae) alimentado com capim-elefante em diferentes temperaturas. *Rev. Brasil. Entomol.* **53**, 614-619.
- Paital, B. (2013). Antioxidant and oxidative stress parameters in brain of *Heteropneustes fossilis* under air exposure condition; Role of mitochondrial electron transport chain. *Ecotoxicol. Environ. Saf.* **95**, 69-77.
- Paital, B. (2014). Modulation of redox regulatory molecules and

- electron transport chain activity in muscle of air breathing fish *Heteropneustes fossilis* under air exposure stress. *J. Comp. Physiol. B*, **184**, 65–76.
- Paital, B., and Chainy, G.B.N. (2010). Antioxidant defenses and oxidative stress parameters in tissues of mud crab (*Scylla serrata*) with reference to changing salinity. *Comp. Biochem. Physiol. C*, **151**: 142-151.
- Paital, B., and Chainy, G.B.N. (2012). Biology and conservation of the genus *Scylla* in India subcontinent. *J. Environ. Biol.* **33**, 871-879.
- Paital, B., and Chainy, G.B.N. (2014). Effects of temperature on complex I and II mediated mitochondrial respiration, ROS generation and oxidative stress status in gills of the mud crab *Scylla serrata*. *J. Therm. Biol.* **41**, 104–111.
- Paital, B., and Samanta, L. (2013). A comparative study of hepatic mitochondrial oxygen consumption in four vertebrates by using Clark-type electrode. *Acta Biol. Hung.* **64**, 152–160.
- Paital, B., Kumar, S., Farmer, R., Tripathy, N.K., and Chainy, G.B.N. (2011). In silico prediction and characterization of 3D structure and binding properties of catalase from the commercially important crab, *Scylla serrata*. *Interdiscip. Sci. Comput. Life Sci.* **3**, 1913-2751.
- Paital, B., Kumar, S., Farmer, R., Tripathy, N.K., and Chainy, G.B.N. (2013). In silico prediction of 3D structure of superoxide dismutase of *Scylla serrata* and its binding properties with inhibitors. *Interdiscip. Sci. Comput. Life Sci.* **5**, 69-76.
- Paital, B., Sablok, G., Kumar, S., Singh, S.K., Chainy, G.B.N. (2015). Investigating the conformational structure and potential site interactions of SOD inhibitors on Ec-SOD in marine mud crab *Scylla serrata*: A molecular modeling approach. *Interdiscip. Sci. Comput. Life Sci.* **7**, 1-7.
- Passos, J.F., and Zglinicki, T.V. (2006). Oxygen free radicals in cell senescence: are they signal transducers? *Free. Radic. Res.* **40**, 1277-83.
- Pillay, C.S., Hofmeyr, J.H., Olivier, B.G., Snoep, J.L., and Rohwer, J.M. (2009). Enzymes or redox couples? The kinetics of thioredoxin and glutaredoxin reactions in a systems biology context. *Biochem. J.* **417**, 269–275.
- Radovanovic, T.B., Mitic, S.S.B., Perendija, B.R., Despotovic, S.G., Pavlovic, S.Z., Cakic, P.D., and Saicic, Z.S. (2010). Superoxide dismutase and catalase activities in the liver and muscle of barbel (*Barbus barbus*) from the Danube river, Serbia. *Arch. Biol. Sci. Belgrade*, **62**, 97-105.
- Rhee S., Chae, H., and Kim, K. (2005). Peroxiredoxins: a historical overview and speculative preview of novel mechanisms and emerging concepts in cell signaling. *Free Radic. Biol. Med.*, **38**(12), 1543–1552.
- Rhee, S.G., Kang, S.W., Chang, T.S., Jeong, W., and Kim, K. (2001). Peroxiredoxin, a novel family of peroxidases. *IUBMB Lif.* **52**, 35–41.
- Sanchez, W., Piccini, B., Ditcher, J.M., and Porcher, J.M. (2008). Assessment of seasonal variability of biomarkers in three-spined stickleback (*Gasterosteus aculeatus L.*) from a low contaminated stream: Implication for environmental biomonitoring. *Environ. Int.* **34**, 791–798.
- Takano, H., Zou, Y., Hasegawa, H., Akazawa, H., Nagai, T., and Komuro, I. (2003). Oxidative stress-induced signal transduction pathways in cardiac myocytes: involvement of ROS in heart diseases. *Antioxid. Redox. Sign.* **5**, 789-794.
- Wohlschlag, D.E., and Juliano, R. (2003). Seasonal Changes in Bluegill Metabolism. *Limnol Oceanography*, **4**, 195-209.
- Wollman, E.E., d'Auriol, L., Rimsky, L., Shaw, A., Jacquot, J.P., Wingfield, P. and et al. (1988). Cloning and expression of a cDNA for human thioredoxin. *J. Biol. Chem.*, **263** 15506–15512. ological Analyses. W. B. Saunders Co., Philadelphia. 357 pp.